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# Rat B Cell Activation Factor(BAFF) ELISA kit

**96 Tests**
**Catalogue Number: E02B0309**
**Store all reagents at 2-8°C**
**Valid Period: six months**

**For samples:**
**Serum, plasma, cell culture supernatants, body fluid and tissue homogenate**

**FOR RESEARCH USE ONLY!**
**NOT FOR THERAPEUTIC OR DIAGNOSTIC APPLICATIONS!**
**PLEASE READ THROUGH ENTIRE PROCEDURE BEFORE BEGINNING!**

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## 1. INTENDED USE

This BG BAFF ELISA kit is a 1.5 hour solid-phase ELISA designed for the quantitative determination of Mouse BAFF. This ELISA kit for research use only, not for therapeutic or diagnostic applications!

## 2. PRINCIPLE OF THE ASSAY

BAFF ELISA kit applies the competitive enzyme immunoassay technique utilizing a monoclonal anti-BAFF antibody and an BAFF-HRP conjugate. The assay sample and buffer are incubated together with BAFF-HRP conjugate in pre-coated plate for one hour. After the incubation period, the wells are decanted and washed five times. The wells are then incubated with a substrate for HRP enzyme. The product of the enzyme-substrate reaction forms a blue colored complex. Finally, a stop solution is added to stop the reaction, which will then turn the solution yellow. The intensity of color is measured spectrophotometrically at 450nm in a microplate reader. The intensity of the color is inversely proportional to the BAFF concentration since BAFF from samples and BAFF-HRP conjugate compete for the anti-BAFF antibody binding site. Since the number of sites is limited, as more sites are occupied by BAFF from the sample, fewer sites are left to bind BAFF-HRP conjugate. A standard curve is plotted relating the intensity of the color (O.D.) to the concentration of standards. The BAFF concentration in each sample is interpolated from this standard curve.

## 3. MATERIALS

*All reagents provided are stored at 2-8° C. Refer to the expiration date on the label.*

| | MATERIALS | SPECIFICATION | QUANTITY |
|---|---|---|---|
| 1 | MICROTITER PLATE | 96 wells | stripwell |
| 2 | ENZYME CONJUGATE | 6 mL | 1 vial |
| 3 | STANDARD A | 0 pg/mL | 1 vial |
| 4 | STANDARD B | 100 pg/mL | 1 vial |
| 5 | STANDARD C | 250 pg/mL | 1 vial |
| 6 | STANDARD D | 500 pg/mL | 1 vial |
| 7 | STANDARD E | 1000 pg/mL | 1 vial |
| 8 | STANDARD F | 2500 pg/mL | 1 vial |
| 9 | SUBSTRATE A | 6 mL | 1 vial |
| 10 | SUBSTRATE B | 6 mL | 1 vial |
| 11 | STOP SOLUTION | 6 mL | 1 vial |
| 12 | WASH SOLUTION (100 x) | 10 mL | 1 vial |
| 13 | BALANCE SOLUTION | 3 mL | 1 vial |
| 14 | INSTRUCTION | 1 | |

**NOTE:** The BALANCE SOLUTION is used only when the sample is **cell culture** supernatants**, body fluid and tissue homogenate**; if the sample is serum or plasma, then the BALANCE SOLUTION is a superfluous reagent.

| The types of sample: | |
|---|---|
| Sample I: | serum or plasma |
| Sample II: | **cell culture** supernatants**, body fluid and tissue homogenate** |

## 4. SPECIMEN COLLECTION AND STORAGE

**Serum -** Use a serum separator tube and allow samples to clot for 2 hours at room temperature or overnight at 2-8°C. Centrifuge at approximately 1000 × g (or 3000 rpm) for 15 minutes. Remove serum and assay immediately or aliquot and store samples at -20°C or -80°C.

**Plasma -** Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge samples for 15 minutes at 1000 × g (or 3000 rpm) at 2 - 8°C within 30 minutes of collection. Assay immediately or aliquot and store samples at -20°C or -80°C.

**Tissue homogenates** - The preparation of tissue homogenates will vary depending upon tissue type. For this assay, tissues were rinsed in ice-cold PBS (0.02mol/L, pH 7.0-7.2) to remove excess blood thoroughly and weighed before homogenization. Minced the tissues to small pieces and homogenized them in a certain amount of PBS with a glass homogenizer on ice. The resulting suspension was subjected to ultrasonication or to two freeze-thaw cycles to further break the cell membranes. After that, the homogenates were centrifugated for 15 minutes at 1500×g (or 5000 rpm). Remove the supernate and assay immediately or aliquot and store samples at -20°C or

-80°C.

**Cell lysates** - Cells should be lysed according to the following directions.

1) Adherent cells should be detached with trypsin and then collected by centrifugation. Suspension cells can be collected by centrifugation directly.
2) Wash cells three times in PBS.
3) Cells were resuspended in PBS and subjected to ultrasonication for 3 times. Alternatively, freeze cells at -20 °C. Thaw cells with gentle mixing. Repeat the freeze/thaw cycle for 3 times.
4) Centrifuge at 1000×g (or 3000 rpm) for 15 minutes at 2-8 °C to remove cellular debris.
5) Assay immediately or store samples at -20°C or -80°C.

**Cell culture supernatants and other body fluids** - Centrifuge cell culture media at 1000 × g (or 3000 rpm) for 15 minutes to remove debris. Assay immediately or store samples at -20°C or -80°C.

NOTE:

1) Samples should be aliquoted and must be stored at -20°C (less than 3 months) or -80°C (less than 6 months) to avoid loss of bioactivity and contamination. If samples are to be run within 24 hours, they may be stored at 2- 8°C. Avoid repeated freeze-thaw cycles.
2) Prior to assay, the frozen sample should be brought to room temperature slowly and mixed gently.
3) Tissue or cell extraction samples prepared by chemical lysis buffer may cause unexpected ELISA results due to the impacts of certain chemicals.
4) Samples containing a visible precipitate must be clarified prior to use in the assay. Care should be taken to minimize hemolysis. Do not use grossly hemolyzed or lipemic specimens.
5) Do not use heat-treated specimens.

## 5. MATERIALS AND EQUIPMENT REQUIRED BUT NOT SUPPLIED

1) Precision pipettors and disposable tips to deliver 10-1000 µl. A multi-channel pipette is desirable for large assays.
2) 100 mL and 1 liter graduated cylinders.
3) Distilled or deionized water.
4) Tubes to prepare sample dilutions.
5) Absorbent paper.
6) Microplate reader capable of measuring absorbance at 450 nm.
7) Centrifuge capable of 3000 × g.
8) Microplate washer or washing bottle.
9) Incubator (37°C).
10) Data analysis and graphing software.

## 6. SAMPLE PREPARATION

1) BLUEGENE (BG) is only responsible for the kit itself, but not for the samples consumed during the assay. The user should calculate the possible amount of the samples used in the whole test. Please reserve sufficient amount of samples in advance.
2) Please predict the concentration before assaying. If values for these are not within the range of the standard curve, users must determine the optimal sample dilutions for their particular experiments.
3) If the samples are not indicated in the manual, a preliminary experiment to determine the validity of the kit is necessary.
4) Owing to the possibility of mismatching between antigen from other resource and antibody used in our kits (e.g., antibody targets conformational epitope rather than linear epitope), some native or recombinant proteins from other manufacturers may not be recognized by our products.
5) Influenced by the factors including cell viability, cell number and also sampling time, samples from cell culture supernatant may not be detected by the kit.
6) Fresh samples without long time storage are recommended for the test. Otherwise, protein degradation and denaturalization may occur in those samples and finally lead to wrong results.

## 7. REAGENTS PREPARATION

1) Bring all kit components and samples to room temperature (20-25 °C) before use.
2) Samples - Please predict the concentration before assaying. If concentrations are unknown or not within the detection range, a preliminary experiment is recommended to determine the optimal dilution. PBS (pH 7.0-7.2) or 0.9% physiological saline can be used as dilution buffer.
3) Wash Solution - Dilute 10 mL of Wash Solution concentrate (100×) with 990 mL of deionized or distilled water to prepare 1000 mL of Wash Solution (1×). If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. The 1× wash solution is stable for 2 weeks at 2-8°C.
4) Do not dilute the other components which are ready- to-use.

## 8. ASSAY PROCEDURE

Please read Reagents Preparation before starting assay procedure. It is recommended that all Standards and Samples be added in duplicate to the microtiter plate.

1) Secure the desired numbers of coated wells in the holder then add 100 uL of **Standards** or **Samples** to the appropriate well in the antibody pre-coated Microtiter Plate. Add 100 uL of PBS (pH 7.0-7.2) in the blank control well.
2) Dispense 10 uL of **Balance Solution** into 100 uL specimens, mix well. (**NOTE**: This step is required when the sample is cell culture supernatants, body fluid and tissue homogenate; if the sample is serum or plasma, then this step should be skipped.)
3) Add 50 uL of **Conjugate** to each well (NOT blank control well). Mix well. Mixing well in this step is important. Cover and incubate the plate for 1 hour at 37°C.
4) Wash the microtiter plate using one of the specified methods indicated below:

a) Manual Washing: Remove incubation mixture by aspirating contents of the plate into a sink or proper waste container. Fill in each well completely with 1× wash solution, and then aspirate contents of the plate into a sink or proper waste container. Repeat this procedure five times for a total of FIVE washes. After washing, invert plate, and blot dry by hitting the plate onto absorbent paper or paper towels until no moisture appears. Note: Hold the sides of the plate frame firmly when washing the plate to assure that all strips remain securely in frame. Complete removal of liquid at each step is essential to good performance.

b) Automated Washing: Wash plate FIVE times with diluted wash solution (350-400 uL/well/wash) using an auto washer. After washing, dry the plate as above. It is recommended that the washer be set for a soaking time of 10 seconds and shaking time of 5 seconds between each wash.

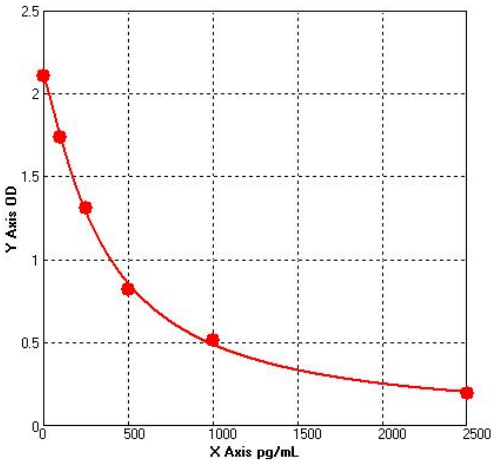
5) Add 50 uL **Substrate A** and 50 uL **Substrate B** to each well including blank control well, subsequently. Cover and incubate for 10-15 minutes at 37°C. (Avoid sunlight).
6) Add 50 uL of **Stop Solution** to each well including blank control well. Mix well.
7) Determine the Optical Density (O.D.) at 450 nm using a microplate reader immediately.

## 9. CALCULATION OF RESULTS

1) The standard curve is used to determine the amount of samples.
2) First, average the duplicate readings for each standard and sample. All O.D. values are subtracted by the mean value of blank control before result interpretation. DO NOT subtract the O.D of standard zero.
3) Construct a standard curve by plotting the average O.D. for each standard on the vertical (Y) axis against the concentration on the horizontal (X) axis, and draw a best fit curve e using graph paper or statistical software to generate a four paramater logistic (4-PL) curve-fit or logit-log linear regression curve. An x-axis

for the optical density and a y-axis for the concentration is also a choice. The data may be linearized by plotting the log of the concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis.

4) Calculate the concentration of samples corresponding to the mean absorbance from the standard curve.
5) Standard curve for demonstration only.




NOTE:

1) Any variation in operator, pipetting and washing technique, incubation time or temperature, and kit age can cause variation in result. Each user should obtain their own standard curve.
2) If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.
3) If specimens generate values higher than the highest standard, dilute the specimens and repeat the assay.

## 10. CERTIFICATE OF ANALYSIS

1) In the same lot CV%: 4.4, 5.6
2) Different lot CV%: 6.6, 7.9
3) Spike Recovery: 94-103%
4) Linearity:

| | Range % |
|---|---|
| 1:2 | 96 – 101 |
| 1:4 | 93 - 107 |
| 1:8 | 92 - 100 |
| 1:16 | 96 - 108 |

5) Sensitivity: The sensitivity in this assay is 1.0 pg/mL.

6) Specificity: This assay has high sensitivity and excellent specificity for detection of BAFF. No significant cross-reactivity or interference between BAFF and analogues was observed. **NOTE**: Limited by current skills and knowledge, it is impossible for us to complete the cross-reactivity detection between BAFF and all the analogues, therefore, cross reaction may still exist in some cases.

## 11. SAFETY NOTES

1) This kit contains small amount of 3, 3', 5, 5'-Tetramethylbenzidine (TMB) in Substrate B. TMB is non-carcinogenic but it is hhazardous in case of skin contact, eye contact, ingestion and inhalation. In case of contact, rinse affected area with plenty of water.
2) The Stop Solution provided with this kit is an acid solution. Wear protective gloves, clothing, and face protection.
3) Care should be taken when handling the Standard because of the known and unknown effects of it.
4) Care should also be taken to avoid contact of skin or eyes with other kit reagents or specimens. In the case of contact, wash immediately with water.
5) Do not pipette by mouth.
6) Avoid generation of aerosols.
7) Waste must be disposed of in accordance with federal, state and local environmental control regulations.
8) All blood components and biological materials should be handled as potentially hazardous. Decontaminate and dispose specimens and all potentially contaminated materials as they could contain infectious agents. The preferred method of decontamination is autoclaving for a minimum of 1 hour a 121.5°C.

## 12. QUALITY CONTROL

1) It is recommended that all standards, controls and samples be run in duplicate. Standards and samples must be assayed at the same time.
2) The coefficient of determination of the standard curve should be $\geq 0.95$ and the highest O.D. should be more than 1.0.
3) Cover or cap all kit components and store at 2-8° C when not in use.
4) Microtiter plates should be allowed to come to room temperature before opening the foil bags. Once the desired number of strips has been removed, immediately reseal the bag with desiccants and store at 2-8°C to maintain plate integrity.
5) Samples should be collected in pyrogen/endotoxin-free tubes.

6) Samples should be frozen if not analyzed shortly after collection. Avoid multiple freeze-thaw cycles of frozen samples. Thaw completely and mix well prior to analysis.
7) When possible, avoid use of badly hemolyzed or lipemic serum. If large amounts of particulate matter are present, centrifuge or filter prior to analysis.
8) When pipetting reagents, maintain a consistent order of addition from well-to-well. This ensures equal incubation times for all wells.
9) Do not mix or interchange different reagent lots from various kit lots.
10) Do not use reagents after the kit expiration date.
11) Read absorbance immediately after adding the stop solution.
12) Incomplete washing will adversely affect the test outcome. All washing must be performed with Wash Solution provided. All residual wash liquid must be drained from the wells by efficient aspiration or by decantation followed by tapping the plate forcefully on absorbent paper. Never insert absorbent paper directly into the wells.
13) Because TMB is light sensitive, avoid prolonged exposure to light. Also avoid contact between TMB and metal, otherwise color may develop.

## 13. CONTACT US

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# 大鼠 B-细胞激活因子 ELISA 试剂盒

应用：定量检测大鼠血清、血浆、细胞液、体液以及组织匀浆中 BAFF 的含量
（本指标为反向竞争法试剂盒，详细实验原理以英文为准）

产品编号：E02B0309

本试剂盒仅供科研使用，不得用于临床及诊断使用！

一． 试剂盒组成

| | 名称 | 规格 | 数量 | 保存 |
|---|---|---|---|---|
| 1 | 预包被平底微孔板 | 96 孔 | 1 可拆卸板 | 2-8℃密封冷藏 |
| 2 | 酶结合物(HRP) | 6 毫升 | 1 瓶 | 2-8℃冷藏 |
| 3 | 标准品 | 1 毫升 | 6 瓶 | 2-8℃冷藏 |
| 4 | 显色剂 A | 6 毫升 | 1 瓶 | 2-8℃冷藏 |
| 5 | 显色剂 B | 6 毫升 | 1 瓶 | 2-8℃避光冷藏 |
| 6 | 终止液 | 6 毫升 | 1 瓶 | 2-8℃冷藏 |
| 7 | 浓缩洗涤液 (100 倍浓缩) | 10 毫升 | 1 瓶 | 2-8℃冷藏 |
| 8 | 平衡液 | 3 毫升 | 1 瓶 | 2-8℃冷藏 |
| 9 | 使用说明书 | | 1 份 | |

注意：平衡液仅仅适用于标本是细胞液、体液以及组织匀浆样品的检测时所使用

二．样本处理及保存：

1. 血清
   采集血清时不加抗凝剂，采集后室温静置 1-2 小时，1000 × g (或 3000 rpm)离心 15 分钟，取上清，-20℃或-80℃分装保存备用。
2. 血浆
   采集血浆，要加入总体积 1%的抗凝剂 (EDTA，肝素等)，采集后先室温或 4℃静置半小时后，1000 × g (或 3000 rpm)离心 15 分钟，取上清，-20℃或-80℃分装保存备用。
3. 组织裂解液
   组织裂解的方式取决于组织的类型，一般来讲，先用预冷的 PBS (0.02mol/L, pH 7.0-7.2) 清洗组织后称重。一般 0.5g 组织加入 500ul 预冷的 PBS (0.02mol/L, pH 7.0-7.2)，在冰上的玻璃容器内研碎。用超声或者反复冻融的方法裂解细胞膜，1500×g (或 5000 rpm) 离心 15 分钟后取上清，-20℃或-80℃分装保存备用。
4. 细胞裂解液

1) 悬浮细胞直接离心取细胞沉淀。贴壁细胞需用胰酶消化后离心取细胞沉淀。检测某些指标时不能用胰酶消化。
2) 用 PBS (0.02mol/L, pH 7.0-7.2) 洗 3 遍。
3) 用少量 PBS 重悬细胞，超声或者反复冻融的方法裂解细胞膜，1500×g (或 5000 rpm) 离心 15 分钟后取上清，-20℃或-80℃分装保存备用。

5. 细胞上清及体液

1000 × g (或 3000 rpm)离心 15 分钟，取上清，-20℃或-80℃分装保存备用。

注意事项：

1. 样本收集完毕后，要分装保存在-20°C (少于 3 个月) 或-80°C (少于 6 个月)以保持蛋白活性和避免污染。避免反复冻融 如果要在 24 小时内分析样本，可以保存在 2- 8°C。
2. 冻存的样本应该缓慢的恢复到室温，然后慢慢地混匀，不能加热来融化样本。
3. 某些化学裂解液会对某些指标的检测会有干扰，比如 SDS，Triton。谨慎使用。
4. 样本液中含有沉淀物会对 ELISA 有干扰，务必离心去除。
5. 不要使用高脂血或溶血的样本，对 ELISA 有干扰，导致检测结果不准确。
6. 其他样本的准备方法，请联系蓝基技术部售后服务人员。

三．试剂准备

1. 试剂准备：所有试剂都必须在使用前达到室温，使用后请立即按照说明书要求保存。
2. 洗液的配制：按 1:100 的比例配制洗液备用。

四．操作步骤（实验前必须仔细看实验准备）

1. 取出试剂盒，于室温（20-25℃）放置 30 分钟。
2. 取出酶标板，按照标准品的次序分别加入 100 μL 的标准品溶液于空白微孔中。
3. 空白微孔中加入 100 μL 的样品，空白对照加入 100 μL 的蒸馏水。
4. 加 10 μL 的平衡液于已加样品的孔中（不含空白对照孔），如果标本是血清或者血浆，此步骤忽略。
5. 在各孔中加入 50 μL 的酶标记溶液（不含空白对照孔）。
6. 将酶标板用封口胶密封后，37℃孵育反应 1 小时（在孵育箱中保持稳定的温度与湿度）。
7. 充分清洗酶标板 5 次，保持各孔有充足的水压（浓缩洗涤液以 1：100 的比例与蒸馏水稀释）。
8. 酶标板洗涤后用吸水纸彻底拍干。
9. 各孔加入显色剂 A 50 μL。
10. 各孔加入显色剂 B 50 μL。
11. 37℃下避光反应 10-15 分钟。
12. 各孔加入 50 μL 终止液，终止反应。
13. 将微孔板至于 450nm 波长的酶标仪下进行读值（吸光度值即为 OD 值）。

五．实验注意事项

1. 实验操作中必须使用一次性吸头，避免交叉污染。
2. 加样：加样时，要控制加样速度，避免第一孔与最后一孔之间的时间间隔过大，否则将会导致不同的预孵育时间，从而影响实验的准确性以及重复性；一般加样时间控制在 10 分钟内，如果样本数量过多，可使用多道移液器。
3. 孵育：样品要在密闭的容器内进行孵育，严格按照说明书上规定的孵育时间和温度进行。
4. 洗涤：洗涤过程中反应孔中的残留的洗涤液应在滤纸上充分拍干，同时要消除板底残留的液体和手指痕迹，避免影响最后的酶标仪读数。
5. 反应时间的控制：加入底物后请定时观察反应孔的颜色变化（比如，10 分钟左右），如果颜色较深，请提前加入终止液终止反应。
6. 建议实验前预测样品含量，如样品浓度过高，应对样品进行稀释，计算结果时乘以相应

的稀释倍数。

7. 建议使用本试剂盒时先做预实验（即先做标准曲线，试用几个标本），如果对本试剂盒有任何疑问，可和所购经销商联系，如果因运输过程导致试剂盒失效，可要求调换，但概不承担产品本身以外的任何损失。